THE STRUCTURES OF PLURACIDOMYCINS, NEW CARBAPENEM ANTIBIOTICS

Sir:

New β -lactam antibiotics which have broad antibacterial and strong β -lactamase inhibitory activities were detected among the metabolites of a new strain of streptomycete named *Streptomyces pluracidomyceticus* nov. sp. (PA-41746). On paper electrophoresis at pH 7.0, certain of these components migrated toward the anode with greater mobility than that of olivanic acids^{1,2)} (Table 1). From these preliminary experiments we assumed that these components should be new carbapenem compounds presumably having two or three acidic functions, and named them pluracidomycins (PLM).

Pluracidomycins were extracted from the fermentation filtrate using a phase-transfer reagent, separated by ion-exchange resin to factors A, B and C, and purified by reverse-phase chromatography and gel filtration in the forms of sodium salts. In a typical batch, 200 liters of the filtrate gave about 450 mg of factor A, 100 mg of factor B and 150 mg of factor C. Pluracidomycins are fairly stable in the pH range of $6.0 \sim 6.5$. Their UV and ¹H NMR spectra are shown in Figs. 1 and 2, respectively. Besides pluracidomycins, penicillin N, cephamycin C, deacetoxycephalosporin C, epithienamycin A and MM17880 were detected in the broth, and also the sulfoxides of epithienarnycin B and D which were characterized as new compounds.

From the ¹H NMR spectra it was easily pointed out that the pluracidomycins all have the sulfated olivanic acid structure^{1~8)}. However, they lack the *N*-acetyl proton signal found in olivanic acids such as MM4550 (4), MM13902 (5) and MM-17880 (6) and have neither -CH=CH- nor $-CH_2$ - CH_2 - functions. Therefore, pluracidomycins can be distinguished from the sulfated olivanic

Fig. 1. UV spectra of pluracidomycins in H₂O.



acids in the substituents at C-3. This conclusion was also confirmed by the ¹³C NMR data listed in Table 2.

The main component, pluracidomycin A (PLM A) shows the highest mobility on paper electrophoresis (Table 1) and has no ¹H and ¹³C signals attributable to the C-3 substituent, which may be an acidic function. The IR spectrum of PLM A exhibited broader and stronger absorption bands at $1200 \sim 1260 \text{ cm}^{-1}$ and $1030 \sim 1080 \text{ cm}^{-1}$ than those of MM4550, therefore, the substituent at C-3 was concluded to be a sulfonate as shown in the structure (1), which was also agreeable with the elemental analysis. PLM A is probably identical with antibiotic SF 2103A found by the Meiji-Seika group⁴.

Pluracidomycin B (PLM B) as well as pluracidomycin C (PLM C) has ¹H signals essentially

Table 1. Paper electrophoresis (1/30 M phosphate buffer pH 7.0; 12 V/cm; 1.5 hours).

Antibiotics	Relative mobility
Epithienamycin A	1.0
MM4550	1.9
MM17880	1.9
PLM A	3.0
PLM B	2.7
PLM C	1.9

Table 2. ¹⁸C NMR in D₂O^a.

Carbon No.	PLM A ^b	PLM B°	PLM C°
2	139.2 (s)	140.0 (s)	140.0 (s)
3	129.0 (s)	140.6 (s)	140.5 (s)
4	32.8 (t)	30.5 (t)	30.4 (t)
5	55.1 (d)	54.7 (d)	54.7 (d)
6	58.7 (d)	59.1 (d)	59.2 (d)
7	168.7 (s)	166.5 (s)	166.4 (s)
8	73.7 (d)	74.0 (d)	73.9 (d)
2-COONa	177.9 (s)	177.5 (s)	177.5 (s)
8-CH ₃	19.4 (q)	19.3 (q)	19.3 (q)
$S(O)-CH_2-$		60.2 (t)	
S(O)-CH-			
OH	-		86.5 (d)
-COONa	_	172.3 (s)	

^a δ ppm from DSS (internal CH₃CN, $\delta = 1.7$).

^b Recorded with a Varian XL-100-12A spectrometer.

Recorded with a Varian XL-200 spectrometer.



Fig. 3. Structures of pluracidomycins and related compounds.



identical with those of MM4550¹⁻³⁾ except the signals due to the substituents at C-3. Moreover, since PLM B and C exhibit the ¹³C chemical shifts of C-3 almost identical to that of MM- $4550^{1,3)}$, both compounds should also have the sulfoxide function adjacent to the C-3 substituent as in MM4550.

Different from MM4550, PLM B has a methylene group (δ ¹H; 4.31, 2H, s., δ ¹³C; 60.2 t.) and a carbonyl function (δ ¹³C; 172.3 s.) in the C-3 substituent. Taking account of the paper electrophoresis and the elemental analysis the carbonyl group is probably a constituent of a carboxyl function. PLM B has much stronger IR bands assignable as COO⁻ at 1600 cm⁻¹ and 1400 cm⁻¹ in comparison with those of PLM A and C or MM4550. This evidence suggests that PLM B has the structure (2) as shown in Fig. 3.

PLM C has a methine group (δ^{1} H; 5.86, 1H, s., δ^{13} C; 86.5, d.) next to the sulfoxide on C-3, and no additional ¹³C signals could be observed. From the ¹H and ¹³C chemical shifts the presence of at least one hydroxyl group, on this methine is likely. The ¹H NMR of PLM C in DMSO-*d*₆ dramatically changed this methine signal from 5.86 ppm to 9.62 ppm (1 H, broad singlet. inextinguishable by D₂O-addition), but the ¹H NMR of the recovered sample in D₂O regenerated the original methine signal at 5.86 ppm. This change

suggests the conversion,
$$-S-CH\langle OH \rightleftharpoons -S-CHO, OH \rightleftharpoons -S-CHO, OH \rightleftharpoons -S-CHO, OH \rightleftharpoons -S-CHO, OH \circlearrowright$$

and the structure of PLM C is concluded to be (3) or (3'). Among pluracidomycins, only PLM C was positive to 2,4-dinitrophenyl hydrazine and the formation of hydrazone or semicarbazone was confirmed by HPLC. Further, PLM C was reduced by sodium borohydride to the alcohol (7), which also has strong β -lactamase inhibitory activity.

The relative configurations of the C-5, C-6, C-8 and the sulfoxide of PLM B and C should be identical with those of MM4550 because of the good correlation of ¹H chemical shifts and coupling constants¹⁻⁸⁾. MM4550 has been related biosynthetically to MM22380⁵⁾ (epithienamycin A), the stereochemistry of which was elucidated as 5-*R*, 6-*R*, 8-*S*⁶⁾. With respect to the sulfoxide of MM4550, the configuration is inferred to be *R* from the CD spectrum¹⁾ which closely resembles that of C-19393⁷⁾ (carpetimycin) whose structure has been established by X-ray analysis^{8,9)}.

As the chromophores of pluracidomycins and MM4550 are different, the direct comparison of

Table 3. Antibacterial activity of pluracidomycins A, B and C.

	MIC (µg/ml)ª		
Organism	PLM A	PLM B	PLM C
Staphylococcus aureus 209P JC-1	25	50	25
Staphylococcus aureus C-14 ^b	50	50	50
Streptococcus pyogenes C-203	>50	50	50
Escherichia coli NIHJ JC-2	6.3	6.3	12.5
Escherichia coli 377°	6.3	6.3	6.3
Klebsiella pneumoniae SRL-1	6.3	12.5	25
Klebsiella sp. 363b	6.3	6.3	6.3
Proteus mirabilis PR-4	12.5	25	12.5
Proteus vulgaris CN-329	25	50	50
Enterobacter cloacae 233	12.5	100	>100
Serratia marcescens ATCC 13880	12.5	25	100
Pseudomonas aeruginosa ATCC 25619	>100	>100	>100

^a Determined by agar dilution method in sensitivity-disc agar and inoculated by one loopful of *ca*. 10⁶ cells per ml.

^b Penicillinase producing strain.

^c Cephalosporinase producing strain.

Source of	Closeb	Minimum effective concentration $(\mu g/ml)^{c}$		
β-lactamase ^a	Class	PLM A	PLM B	PLM C
Escherichia coli 6	Ib	0.008	0.008	0.004
Enterobacter cloacae 92	Ia	0.001	0.008	0.008
Proteus vulgaris 31	Ic	<0.001	<0.001	<0.001
<i>Escherichia coli</i> W3110 RTEM	IIIa	0.25	0.002	0.001
Klebsiella sp.363	IV	0.008	0.008	0.004
Enterobacter cloacae 53	IVa	0.125	<0.001	<0.001

Table 4. Inhibition of β -lactamases produced by Gram-negative bacteria by pluracidomycins.

- ^a Enzyme preparations used were partially purified.
- ^b Classification of RICHMOND and SYKES.
- Inhibitor was incubated with enzyme at room temperature for 10 minutes prior to adding nitrocefin (50 μg/ml) and minimum concentration to inhibit color change was determined.

their COTTON-effects is of no use for the determination of the absolute configuration. Therefore, PLM B was reduced to the corresponding sulfide (8) by TiCl_{8}^{10} , and the CD spectrum was compared with that of epithienamycin A. Both spectra showed the same COTTON-effects, accordingly, pluracidomycins should have the configuration identical with that of MM4550 as shown in Fig. 3. The sulfoxides of PLM B and C are arranged spatially in the same direction as those of MM4550, carpetimycins and asparenomycins¹⁰⁾, but in the case of PLM C only the sulfoxide should be designated by the *S*-configuration according to the sequence rule.

The biological properties of pluracidomycins are summarized in Tables 3 and 4*. The details will be reported separately.

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^{*} The *in vitro* data were obtained by Dr. T. YOSHIDA and his colleague.

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